

# **On the question of activity of oxidative branch of Pentose Phosphate shunt in *pgl* mutant of *E. coli***

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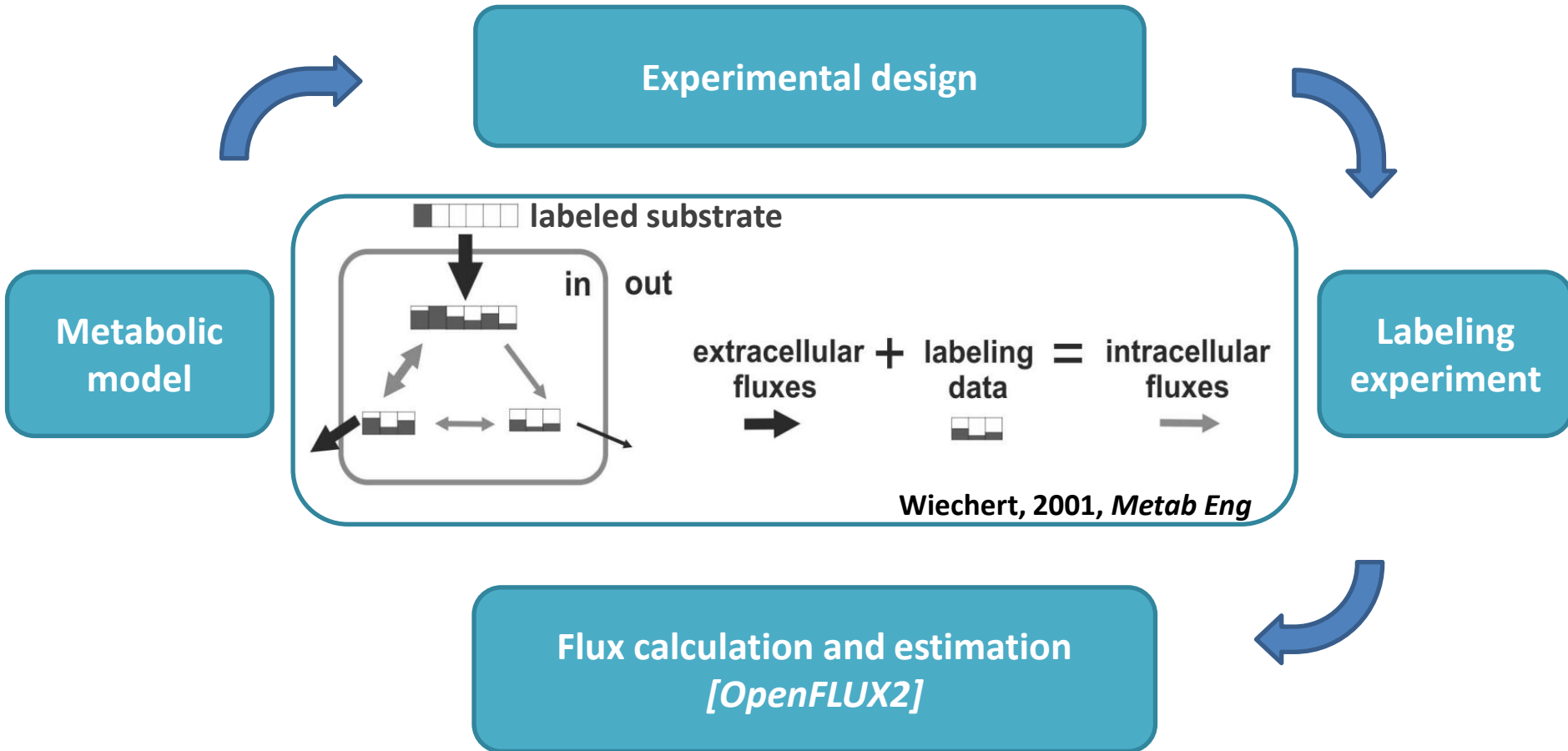
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**July 4–8, 2022  
Novosibirsk**

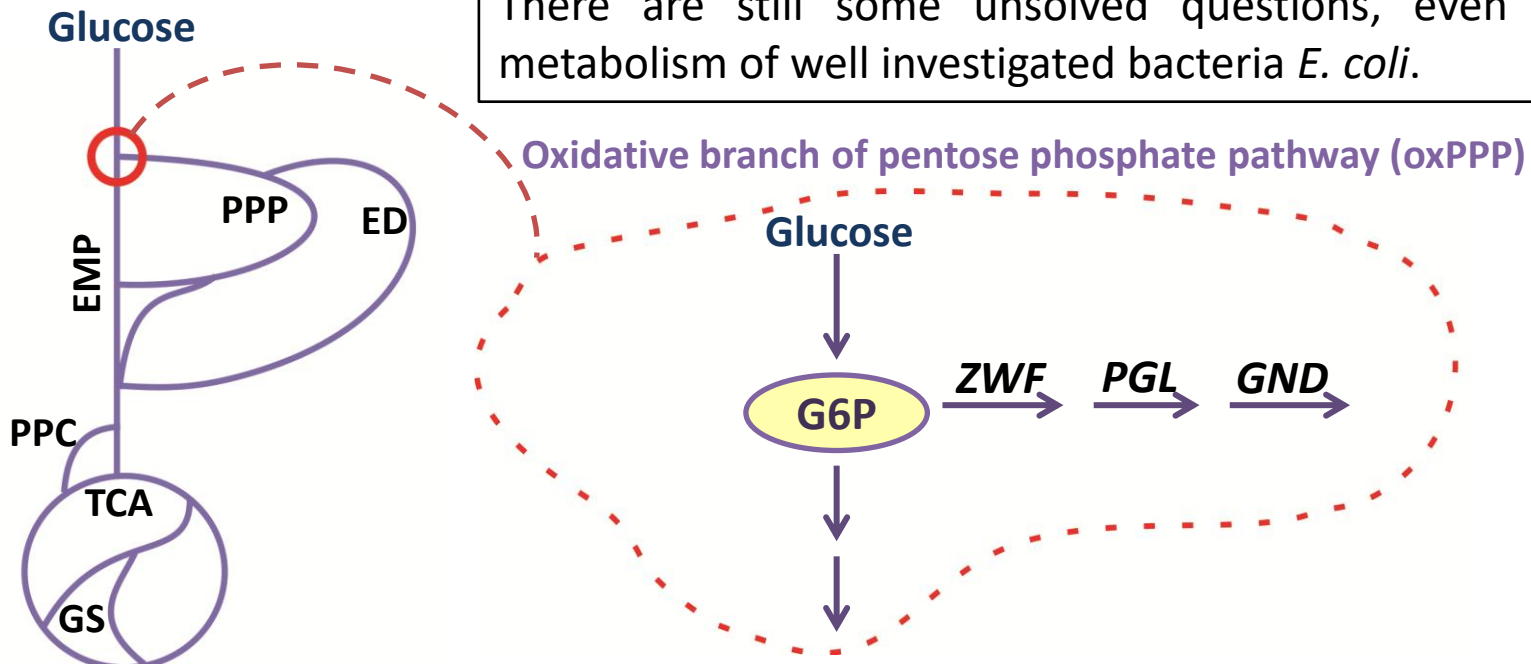
# The principle scheme of $^{13}\text{C}$ -MFA ( $^{13}\text{C}$ metabolic flux analysis)

$^{13}\text{C}$  metabolic flux analysis ( $^{13}\text{C}$ -MFA) is complex method for metabolism investigations allowing quantitative estimation of actual carbon metabolic fluxes distribution *in vivo*.



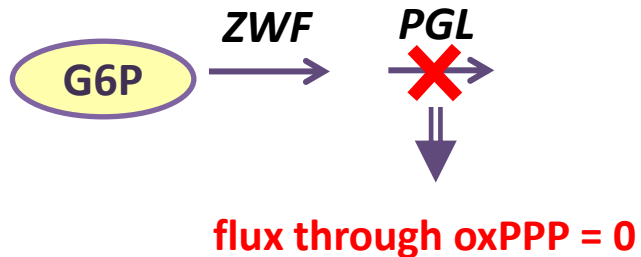
# The role of PGL reaction in *E. coli* central metabolism

There are still some unsolved questions, even about central metabolism of well investigated bacteria *E. coli*.

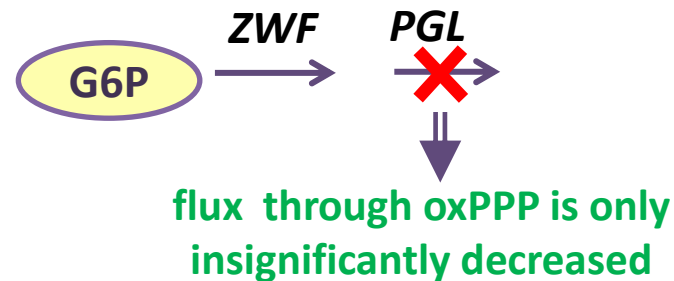


## Flux distribution in *E. coli* BL21 or MG1655 $\Delta pgl$

Flux balance analysis (FBA)\*



$^{13}\text{C}$  metabolic flux analysis\*\*



What's the true  
?

\*Juan C. Aon et al., 2008, *Appl Environ Microbiol*

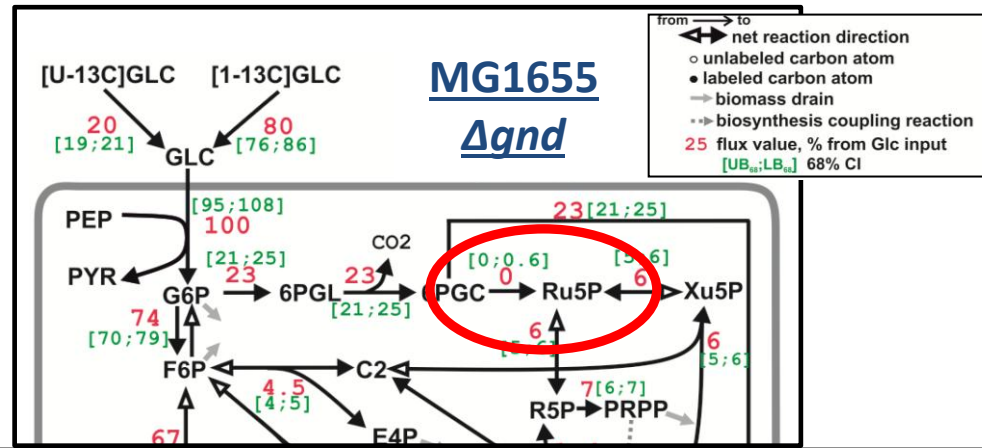
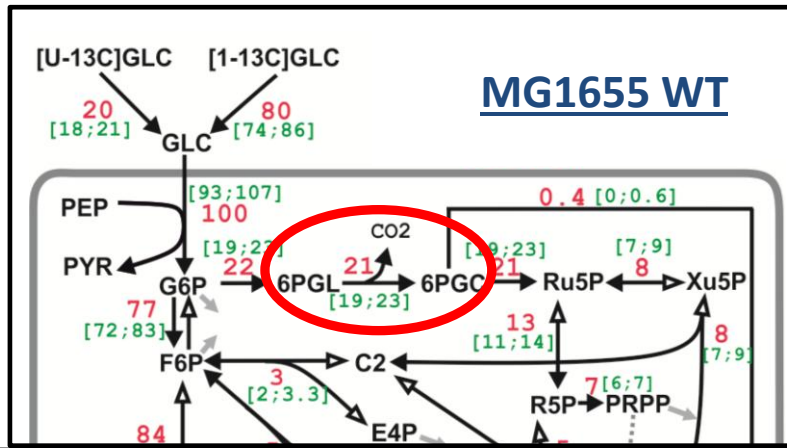
\*Jonathan M. Monk et al., 2016, *Cell Syst*

\*\* C. P. Long and M. R. Antoniewicz, 2019, *Metab Eng*

\*\* Christopher P Long et al., 2017, *Metab Eng*

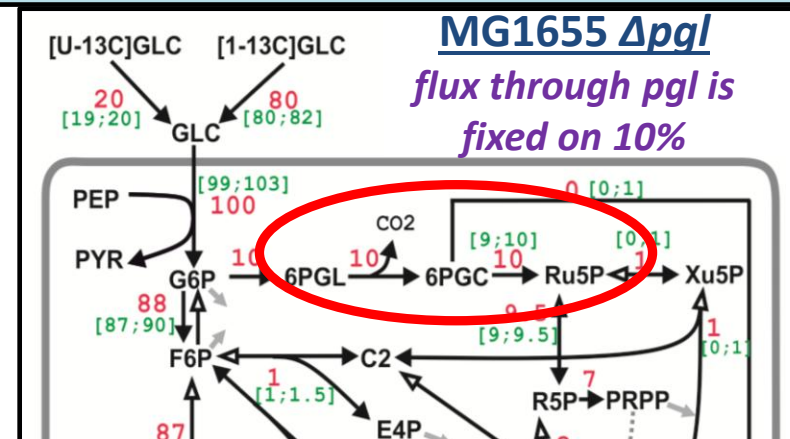
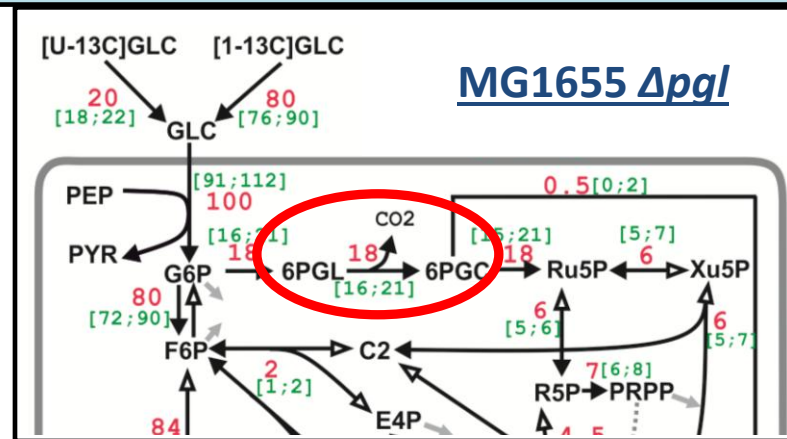
\*\* this work

# <sup>13</sup>C-MFA of *E. coli* oxPPP mutants



from → to  
 ⇌ net reaction direction  
 ○ unlabeled carbon atom  
 ● labeled carbon atom  
 → biomass drain  
 ⇨ biosynthesis coupling reaction  
 25 flux value, % from Glc input  
 [UB<sub>in</sub>;LB<sub>in</sub>] 68% CI

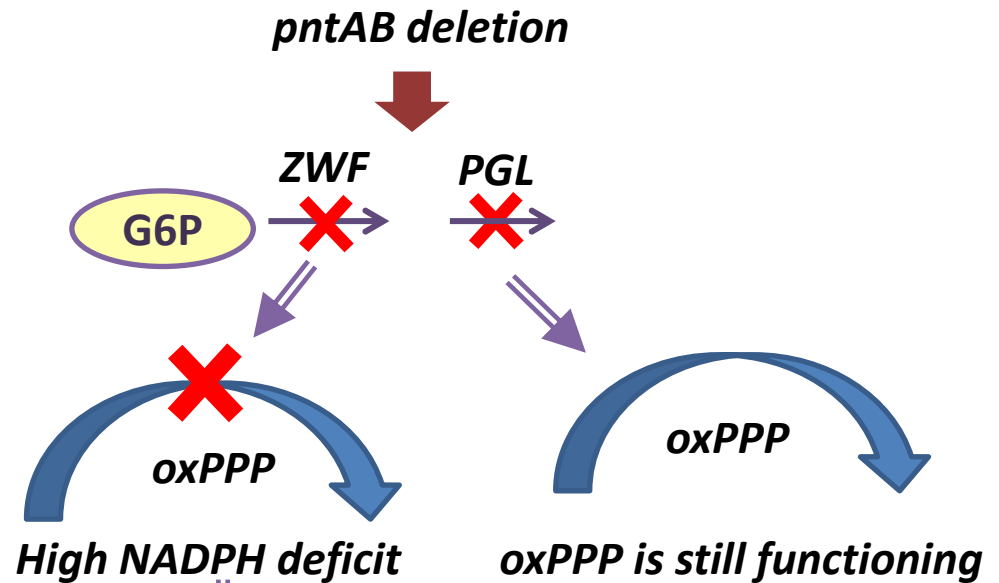
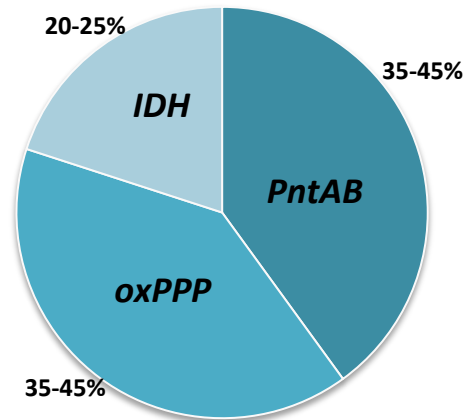
The flux solution with the best accordance with experimental data for  $\Delta pgl$  mutant corresponds to carbon flux through the oxPPP of **18%** of input glucose in contrast with **25%** in parent strain. In flux calculations minimal value of SSR is commonly used as conditional criterion. Formally, any flux distribution providing SSR value which passes  $\chi^2$ -criterion can be recognized as acceptable. However, flux calculation under assumption that *pgl* inactivation blocks oxPPP led to only statistically unacceptable solution. We determined that flux through PGL reaction responded to  $\chi^2$ -criterion on minimal 10% level. It means that the the flux through the oxPPP decreases by no more than 2.5-times in the  $\Delta pgl$  mutant.



## Role of oxPPP in NADPH supplementation

According to Sauer et al.\*, the main sources of NADPH in *E. coli* are oxidative pentose phosphate pathway (oxPPP), transhydrogenase PntAB and isocitrate dehydrogenase (IDH).

Reactions contribution to production of NADPH required for biosynthesis



**Dramatically decreased growth**

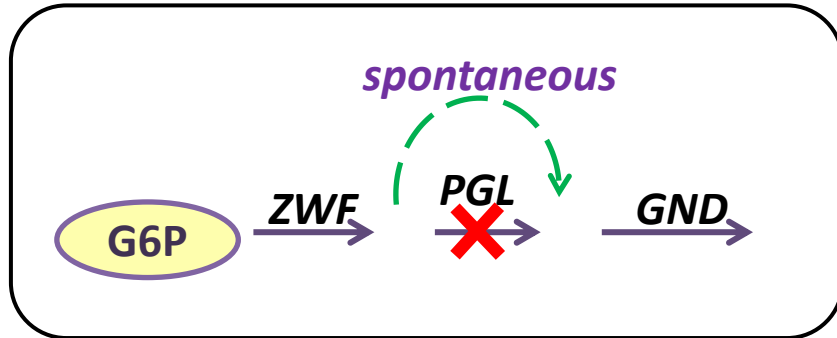
The  $\Delta pgl\Delta pntAB$  and  $\Delta gnd\Delta pntAB$  mutants demonstrated decrease of growth rate in 1.3 and 1.7 times, respectively. This additionally confirms that *pgl* inactivation do not abolish activity of oxPPP as in case of *zwf* inactivation. The interesting fact that mutant  $\Delta gnd\Delta pntAB$  grows 1.3 times slower than  $\Delta pgl\Delta pntAB$  mutant despite of activity of Entner-Doudoroff bypass, indicating, probably, that the first strain has more restriction in NADPH supplementation.

Strain	Growth rate, h <sup>-1</sup>
MG1655	0.63
MG1655 $\Delta zwf$	0.55
MG1655 $\Delta zwf \Delta pntAB$	0.09
MG1655 $\Delta pgl$	0.48
MG1655 $\Delta pgl \Delta pntAB$	0.38
MG1655 $\Delta gnd$	0.56
MG1655 $\Delta gnd \Delta pntAB$	0.33

\*U. Sauer et al., 2004, *J Biol Chem*

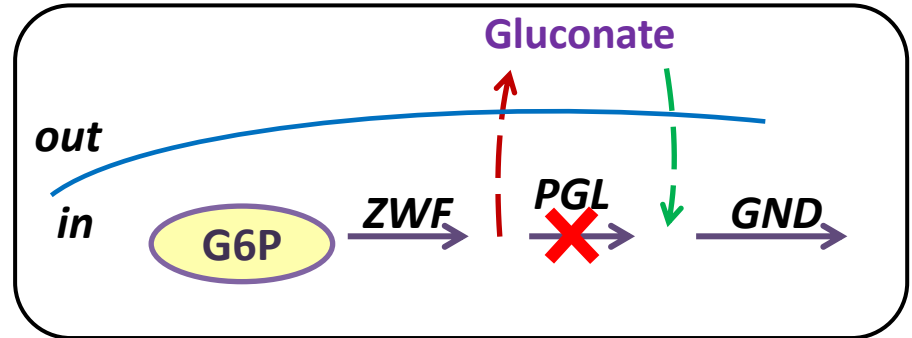
# Role of gluconate bypass in oxPPP fluxes maintaining

## Possible ways of oxPPP functioning in $\Delta pgl$ mutant



**Full compensation by spontaneous hydrolysis of 6-phosphogluconolactone.**

**It looks doubtful**



**Bypass through gluconate**

Testing of hypothesis

We analyzed the growth of mutant with inactivation of gluconate utilization genes, *gntT*, *gntKU* and *gntP*, on minimal medium with glucose. Simultaneous inactivation of these genes reduces growth of  $\Delta pgl$  and  $\Delta pgl\Delta pntAB$  mutants on glucose. These observations allow us to suppose that in MG1655 $\Delta pgl$  strain the PGL reaction can be at least partly bypassed by the dephosphorylation of the substrate of PGL reaction, 6-phosphogluconolactone, and its conversion to gluconate, which then is consumed by the cells.